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HIV Protease Inhibitor HOE/BAY 793, Structure–Activity Relationships in a Series of C₂-Symmetric Diols

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Abstract—A detailed structure-activity relationship of C_2 -symmetric diol inhibitors of HIV-1 protease leads to inhibitor 6 (HOE/BAY 793) which is outstanding in the inhibition of the enzyme and in the inhibition of viral replication in HIV infected cell culture (IC₅₀: 0.3 nM; EC₅₀: 3 nM). There are well defined steric requirements for the design of the side chains P1-P3 of the inhibitors. In addition, all three side chains need to be lipophilic. While the enzyme tolerates hydrophilic substituents in some cases, drastic reductions in anti-HIV activity are observed in cell culture, most likely due to insufficient cell penetration.

Introduction

The rapid and global spread of the acquired immune deficiency syndrome (AIDS) pandemic has initiated intensive efforts to treat this aggressive disease. The etiological cause of AIDS is the human immunodeficiency virus (HIV), a retrovirus that belongs to the group of lentiviruses. By analogy to the processing of proteins of other retroviruses, some HIV proteins are first translated into long precursor proteins, which later have to be cleaved to the active proteins by a virus encoded protease. This specific processing of the HIV gag and gag pol polyprotein gene products by HIV protease is essential for viral maturation. Therefore, inhibition of HIV protease is considered a promising target for antiviral chemotherapy.¹

The HIV protease belongs to the aspartic protease family and therefore the design of inhibitors was influenced by the knowledge obtained in the study of renin and other related enzymes. The most potent inhibitors reported so far are pseudopeptidic compounds containing transition-state inserts instead of the cleavage sites in the natural or unnatural peptide substrates.² Recently several non-peptide inhibitors of the HIV-protease have been reported.³

While the active center of typical aspartic proteases is located on a single protein chain, the HIV protease is enzymatically active only upon association to a C₂ symmetric dimer.⁴ This prompted us⁵ to synthesize C₂ symmetric inhibitors. In previous reports we described a series of inhibitors with a central phosphinic acid or a hydroxymethylphosphinic acid unit in a C₂ symmetric peptidic environment.⁶

In this article we concentrate on C₂-symmetric compounds with a vicinal diol group as a central unit.

The same approach has been concurrently followed by Kempf et al.⁷ In this paper we present detailed structure—activity relationships (SARs) both for enzyme inhibition and for in vitro activity. Interestingly the two SARs are not at all parallel: while good enzyme inhibition is an absolute requirement, it does not necessarily result in good antiviral in vitro activity. The systematic variation of substituents of P1, P2 and P3 led to the inhibitor HOE/BAY 793 (compound 6) with outstanding potency, especially in HIV infected cell culture.

Chemistry

The synthesis of the central building blocks and inhibitors is outlined in Scheme 1 which shows the synthesis of compound 6 as a representative example. Methods similar to those shown in Scheme 1 have also been reported by several groups.7-9 The synthesis of the symmetric diamino diol moiety 3 was achieved in two ways. Route A starts from the N-protected phenylalaninal 1 which is converted into 3 by reductive pinacol coupling. When samarium(II) iodide is used as reducing agent the three isomers SRRS, SRSS, SSSS are obtained in the ratio 2.3:1:1.4. For the stereospecific synthesis of the pure SRRS isomer [V₂Cl₃(THF)₆]₂[Zn₂Cl₆] is used, which has been shown⁸ to yield the isomers in a 6:1.5:1 ratio. Kammermeier et al. 86 describe a suitable synthesis for the large scale preparation of 6. Route B starts from the D-mannitol based bisepoxide 2 which is reacted with (C₆H₅)₂CuLi and the resulting free hydroxyl groups are subsequently converted into amino groups. This chiral pool derived route has the advantage of a correctly preformed stereochemistry.

The central building blocks of the inhibitors 12, 13 and 17 were obtained by route A, those of 18–22 by route B.

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Scheme 1. Representative synthesis of inhibitors. (a) SmI_2 or $[V_2Cl_3(THF)_6]_2[Zl_2Cl_6]$; (b) HCl/MeOH or $H_2/Pd/C$; (c) $(C_6H_5)_2CuLi$; (d) $p-O_2NC_6H_4SO_2Cl$; (e) NaN_3/DMF ; (f) H_2/Pd ; (g) HCl/MeOH; (h) 1. Z-Val-OH, TOTU, (i) $H_2/Pd/C$; (k) DCC, HOOBt.

Active ester coupling of 4 and the terminal unit 5¹⁰ with DCC and HOOBT yielded inhibitor 6. The synthesis of inhibitors 14 and 15 (Scheme 2) is achieved by conversion of the N-Boc protected 3 into the cyclic sulfite 7 and subsequent ring opening by azide. Compound 15 is obtained from 14 by catalytic hydrogenation.

Compounds 10, 11, 16 and 23-51 were obtained in analogy to 6 by application of various suitable protecting groups.

Results and Discussion

Tables 1–5 give a representative selection of the inhibitors that were evaluated for their ability to inhibit HIV-protease and in the HIV infected cell culture. Values for inhibition of the enzyme (IC₅₀ values) were determined by a peptide hydrolysis assay that employs the synthetic heptapeptide substrate H₂N-Ser-Phe-Asn-Phe-Opr-Gln-Ile-OH, where Opr stands for 5-oxaproline, and quantitation via HPLC.¹¹ The EC₅₀ values represent the ability of the compound to inhibit the replication of the virus by 50% based on the cytopathic effect of HIV-1^{12a} or on reduction of viral antigen in the supernatant of infected human T-cells.^{12b}

Transition state mimic

Tables 1 and 2 demonstrate the influence of different groups in the center of this type of inhibitor. As expected, hydroxyl groups strongly enhance the inhibition of the enzyme. Compound 11 containing the bis-hydroxyl moiety shows fifty-fold better activity compared to the hydrocarbon-centered inhibitor 10. This reflects the binding contribution of the diol transition state mimic. Nevertheless some activity is achieved even without hydroxyl groups, provided that other parts of the inhibitor (in this case P2 and P3) provide enough binding energy. Although 11 is a very good inhibitor of the enzyme, it is only slightly active in the cell culture assay. The main reason seems to be the missing lipophilic substituent in P1, which is needed for cell penetration.

Contrary to our expectations the configuration of the hydroxyl groups has not much impact on the interaction of the inhibitor with HIV protease as can be seen from Table 2. Compound 6 (RR-configuration) for example is only somewhat more active than its isomers 12 and 13 (SS- and RS-configurations). However, a strong influence of the hydroxyl group configuration on the HIV inhibitory activity in cell culture is observed. Compound 6 is about thirty times more active than the unsymmetrical isomer 13. At the moment we do not have an explanation for this result.

Scheme 2. (a) SOCl,; (b) NaN, DMF; (c) 1. HCl/dioxane; (d) 1. Boc-Val-OH, TOTU, Et, N; 2. HCl/dioxane; 3. 5, TOTU, Et, N; (e). H₂/Pd/C.

Interestingly one hydroxyl group is sufficient for both enzyme and cell culture inhibition, as found in 16. Substitution of one hydroxyl group by an amino group as in 15 does not weaken the enzyme inhibitory activity as compared with 13, but leads to a nearly complete loss of activity in the cell culture assay. This could be due to the basic nature of the amino group impairing cell penetration. Substitution by the neutral, but larger azide group in 14 leads to a loss of activity by a factor of 30 already at the enzyme level.

P1-position

Table 3 summarizes data on the influence of side chains filling the P1 position. Medium sized lipophilic side groups (e.g. in 6 and 22) are favourable for enzyme inhibition as well as for activity in cell culture. The best result is obtained with the benzyl group in 6. Elongation of the alkyl side chain by one methylene group in 17 (or possibly 21) dramatically decreases

activity compared to 6. Enlargement of its volume as in 20 also leads to a decrease of activity. While the introduction of hydrophilic substituents in P₁ as the tyrosine side chain in inhibitor 18 still yields fairly good enzyme inhibition, the activity in the cell assay is decreased by a factor of a hundred, probably because of diminished cell penetration. The fact that P1 side chains are more important for activity in cell culture than for simple enzyme inhibition is demonstrated by compound 11 lacking the P1 side chain, which is highly active on the enzyme and almost inactive in cell culture.

P2-position

The best activity in enzyme and cell assay is shown by compounds with small alkyl side chains in P2 (see Table 4) which are branched in the β -position, e.g. 6, 23, 24, 25 or 26. However, branching in the β -position is not an absolute requirement, the propyl substituent in

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27 also leads to excellent inhibition. Again, the introduction of hydrophilic substituents as in 30 and 31 is compatible with good activity in the enzyme assay, but not with in vitro inhibition. Interestingly the enzyme does not tolerate the amide side chain in 29 nor the imidazole side chain in 32. The enzyme also reacts very sensitively to changes in the backbone of the inhibitor: elongation by a methylene group as in 28 or even more dramatically the fixing of the backbone conformation by a proline residue as in 33 leads to a loss in activity.

P3-position

Table 5 shows data on some representative inhibitors that carry optimized residues in the positions P1 (benzyl) and P2 (isopropyl), varying in the P3-position. Compound 40 is almost identical with the inhibitor A-77003, selected for clinical development by Abbott, 7a except for the RS-geometry at the hydroxyl groups.

Table 1. Influence of OH-groups in the central building block on HIV-1 protease inhibition and antiviral activity; DSNP: see Scheme 1

No.	R	IC so[nM] a)	EC 50[nM] b)
10	Н	100	3,000
11	OH	22	10,000

(a) Concentration of test compound at which 50% inhibition of the activity of HIV-1 protease was observed.¹¹; (b) concentration which inhibited 50% of the cytopathic effect of HIV-1 or the production of p24 antigen in lymphocytes¹².

Table 2. Influence of configuration and functionality in the central building block on HIV-1 protease inhibition and antiviral activity.

DSNP: see Scheme 1

No.	R1 (config.)	R2 (config.)	IC 50 [nM]*)	EC 50 [nM] b)
6	OH (R)	OH (R)	0.3	3
12	OH (S)	OH (S)	0.3	20
13	OH (R)	OH(S)	0.5	217
14	OH (R)	$N_3(S)$	60	10 5
15	OH (R)	$NH_2(S)$	2.1	> 105
16°)	OH (S)	H	0.3	17

(a,b) See Table 1; (c) geometry S corresponds to R in the diol case due to CIP-rules.

Table 3. Influence of variations in P1 on HIV-1 protease inhibition and antiviral activity. DSNP: see Scheme 1

No.	R-	IC _{so} [nM] a)	EC _{se} [nM] ^{b)}
6		0.3	3
11	-H	2.2	10000
17		360	n.d.
18	OH	2.9	300
19		0.4	20
20	MeOOMe	18	175
21	N	140	n.d.
22	Me	1.0	7.5

(a,b) see Table 1

Regarding enzyme inhibition Table 5 further shows that the use of structurally very diverse terminal groups like positively or negatively charged residues, e.g. 34, 35 or 36, polyhydroxy or neutral groups, e.g. 42 or 38 result in highly active HIV protease inhibitors. However, regarding antiviral efficacy, only two of the tested residues, 41 and 43, lead to really good inhibitors in the cell culture assay.

The terminus of 43¹⁰ served as a lead structure for more detailed investigations, that are summarized in Table 6. With the sulfonyl group held in place, enzyme inhibition can be obtained with quite different structures. Comparison of 43 and 47 demonstrates that a side chain in P3 is not a requirement for high activity. Only compound 49 obviously carries a terminal chain which is too long for the P3 pocket of the enzyme. For virus inhibition we get the same overall picture as for P1. The lipophilic side chain is useful for enzyme inhibition, but mandatory for viral suppression in the cell culture assay. Every increase of the hydrophilic character as in the pyridyl compound 45, the N-oxide 46 or even the methoxy derivative 44 and the hydroxyethyl derivative 48 has to be paid for with loss of virustatic activity. In a similar way substitution of the tert-butylsulfonyl group by the more polar sulfonamide containing moiety in 51 is unfavorable for activity in cell culture. On the other hand enhancement of lipophilic properties by substitution of the phenyl residue in 43 by a naphthyl residue leads to the extremely potent inhibitor 6 (HOE/BAY 793).

Table 4. Influence of variations in P2 on inhibitory activity. DSNP: see Scheme 1

DSNP X N H OH H X DSNP

No.	X-	IC 50 (nM) a)	EC _{se} (nM) ^{se}
6	-HN II	0.3	3
23	-HN O	0.3	10
24	-HN II	0.5	15
25	-HN	1.7	60
26	-HN II	0.3	7
27	-HN O	0.4	4
28	-HN	50	10,000
29	-HN CONH2	100	10,000
30	-HN OH	0.3	2,000
31	-HN COOH	0.75	> 100.000
32	-HN O	340	> 10,000
33	N-N-N-N-N-N-N-N-N-N-N-N-N-N-N-N-N-N-N-	20,000	n.d.

Table 5. Influence of variations in terminus on inhibitory activity

No.	R-	IC m (nM) a)	EC _{ss} (nM) b)	
34	H ₂ N O	0.85	> 25000	
35	H ₂ N	25	> 25000	
36	HO	1.4	>25000	
37	но	22	100	
38		2	100	
39		1.3	253	
40	Me N N	15	958	
41		38	41	
42.	OH OH OH H O	0.9	> 25000	
43	OH OH OH H Ö	0.3	23	
	-			

(a,b) see Table 1

(a,b) see Table 1

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Table 6. Influence of variations in terminus on inhibitory avtivity

No.	R-	IC a(nM) a)	EC (nM)	
6		0.3	3	
	\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\			
43	00	3.2	25	
	OMe			
44		0.34	80-400	
	× S			
45		0,8	25,000	
46	_\.\.\.\.\.\.\.\.\.\.\.\.\.\.\.\.\.\.\	7	50000	
40	رِبِهِ ان م	,	30000	
<i>a</i>		4	1,000	
47	λ_{s}			
46	но о о	4.8	6,000	
49	Me(CH ₂), S	, > 10000	25000	
50	0.0 N	4	1200	
51		0.8	500	
	Me S N H O			

(a,b) see Table 1

6 (HOE/BAY 793) Figure 1.

Conclusions

The detailed structure-activity relationship presented here does give a clear picture of the requirements for the P1-P3 sidechains in combination with the central C₂-symmetric building block. All three side chains need to be lipophilic. While the enzyme tolerates hydrophilic substituents in some cases, drastic reductions in anti-HIV activity are observed, most likely due to insufficient cell penetration. The best results are obtained with compound 6 (HOE/BAY 793) which is outstanding in both enzyme and *in vitro* virus inhibition.

Experimental

Chemistry

Proton magnetic resonance spectra were recorded on a Bruker AM 270 spectrometer. Chemical shifts were recorded as δ units relative to tetramethylsilane as internal standard. FAB mass spectra were taken on a VG Analytical 11-250J spectrometer, and relevant data are tabulated as m/z. Anhydrous solvents were obtained as follows: methylene chloride, distillation from P2O5 or drying over 4Å molecular sieves; tetrahydrofuran, distillation from lithium aluminium hydride or use of Aldrich Sure-Seal solvent without further purification; dimethylformamide, drying over 4Å molecular sieves. Flash chromatography was performed on silica gel 60 (230-400 mesh, E. Merck). All solvents for chromatography were reagent grade. Thin layer chromatography was performed with E. Merck or Riedel-de Haën silica gel 60 F-254 (0.25 mm) plates and visualized with phosphomolybdic acid or ninhydrin. The following abbreviations are used: EA, ethylacetate; MC methylene chloride; NEM, N-ethylmorpholine; DMF, dimethylformamide; THF, tetrahydrofuran; DCC, dicyclohexylcarbodiimide; HOBT, 1-hydroxybenzotriazole hydrate; HOOBT, 1-hydroxybenzotriazine; TOTU, O-[(ethoxycarbonyl)cyanomethylenamino]-N, N, N', N'tetramethyluronium-tetrafluoroborate. 13

The synthesis of inhibitors 38, 39, 40 and 3S-hydroxy-2S,5S-diamino-1,6-diphenylhexane was carried out as published.⁷

2S,5S-Diamino-1,6-diphenylhexane-3R,4R-diol Route A. (a) tert-Butoxycarbonyl-L-phenylalaninal 1 (17 g) was dissolved in 500 mL of dry THF and the solution was cooled to 0 °C under argon. One liter of 0.1 molar Sml₂ solution in THF was added in the course of about 20 min and the mixture was subsequently stirred at rt for 30 min. It was acidified to pH 1-2 with 0.1 N aqueous HCl. The mixture was diluted with EA and the organic phase was separated and extracted with 0.1 N HCl, 2 times with Na₂S₂O₃ solution and 2 times with water. After drying over MgSO₄, the extract was concentrated and the residue was chromatographed over silica gel (EA:petroleum ether, 1:2). The fraction N,N'-bis-(tert-butoxycarbonyl)-2S,5Scontaining diamino-1,6-diphenylhexane-3R,4R-diol was recrystallized from ethanol/water (yield 1.61 g).

N, N'-bis-(tert-Butoxycarbonyl)-2S,5S-diamino-1,6-diphenylhexane-3S,4S-diol (1.0g) was obtained from the fraction containing the 3S,4S and the 3R,4S isomer by crystallization from MC/isopropyl ether/heptane. The mother liquor was chromatographed on RP18 silica gel to obtain 0.71 g of N,N-bis-(tert-butoxycarbonyl)-2S,5S-diamino-1,6-diphenylhexane-3R,4S-diol (acetonitrile:water 4:6).

 R_f values [silica gel, ethylacetate:hexane, 1:2]: 0.18 (3R,4R), 0.41 (3S,4S), 0.39 (3R,4S), MS (FAB): 501 (M + H)⁺, 401, 345, 327 (all three isomers); ¹H NMR (270 MHz, DMSO- d_6): 3R,4R isomer: 1.30 (s, 18H), 2.54–2.80 (m, 4H), 3.24 (m, 2H), 4.12 (m, 2H), 4.43 (m, 2H), 6.16 (d, 2H), 7.08–7.27 (m, 10H). 3S,4S Isomer: 1.30 (s, 18H), 2.63 (dd, J = 14 Hz, 9 Hz, 2H), 3.04 (dd, J = 14 Hz, 4 Hz, 2H), 3.42 (m, 2H), 3.71 (m, 2H), 4.57 (d, J = 7 Hz, 2H), 6.60 (d, 2H), 7.11–7.29 (m, 10H). 3R,4S Isomer: 1.24 (s, 9H), 1.32 (s, 9H), 2.62–2.83 (m, 4H), 3.27–3.46 (m, 2H), 3.91–4.12 (m, 2H), 4.94 (d, J = 4 Hz, 1H), 4.62 (d, J = 4 Hz, 1H), 6.28 (d, 1H), 6.31 (d, 1H), 7.08–7.32 (m, 10H).

- (b) The 2S,5S-diamino-1,6-diphenylhexane-3,4-diol isomers were obtained from the corresponding N,N-bis-(tert-butoxycarbonyl)-2S,5S-diamino-1,6-diphenylhexane-3,4-diol isomers by treatment with 3 N HCl in methanol for 15 min at 25 °C and were used without further purification.
- 2S, 5S-Diamino-1,6-diphenyl-3,4-O-isopropylidenehexane-3R,4R-diol (3b). Route B. (a) 1,2R-5R,6-Diepoxy-3,4-O-isopropylidene-3R,4R-diol 2^{16} (1.12 g) was added to a solution of 36 mmol of $(C_6H_5)_2$ CuLi in 60 mL of dry ether at -78 °C under argon. The cooling bath was removed and the mixture was allowed to warm to rt, while stirring. EA (250 mL) was added to the mixture and the mixture was extracted 3 times with a mixture of 25% ammonia and ammonium chloride. The EA phase was washed with NaCl solution, dried and concentrated. The residue was purified over silica gel (methylene chloride:ethylacetate, 97:3 to 90:10). 2R,5R-Dihydroxy-1,6-diphenyl-3,4-O-isopropylidene-3R,4R-diol (1.86 g) was obtained. MS (FAB): 343 (M + H)⁺, 327, 285, 267; ¹H NMR (270 MHz, DMSO- d_6): 1.39 (s, 6H), 2.58 (dd,

- J = 13 Hz, 9 Hz, 2H), 3.43 (dd, J = 13 Hz, 3 Hz, 2H), 3.68 (m, 2H), 3.83 (m, 2H), 5.05 (d, J = 6 Hz, 2H), 7.14–7.32 (m, 10H).
- (b) 2R,5R-Dihydroxy-1,6-diphenyl-3,4-O-isopropylidenehexane-3R,4R-diol (5.6 g) was dissolved in 300 mL of chloroform together with 7.9 g of DMAP. p-Nitrobenzenesulfonyl chloride (14.5 g) was added at rt and the mixture was stirred at 50 °C for 3 h. MC was added and the solution was extracted with bicarbonate solution, KHSO₄ solution and NaCl solution. The organic phase was dried and concentrated. 2R,5R-Di-(4-nitrophenylsulfonyloxy)-1,6-diphenyl-3,4-O-isopropylidenehexane-3S,4S-diol (11.8 g) was obtained. MS (FAB): 713 (M + H)⁺, 697, 510; ¹H NMR (270 MHz, DMSO- d_6): 1.42 (s, 6H), 2.87 (dd, J = 15 Hz, 9 Hz, 2H), 3.11 (dd, J = 15 Hz, 3 Hz, 2H), 4.41 (s, 2H), 5.07 (dm, J = 9 Hz, 2H), 6.95-7.11 (m, 10H), 7.73 (d, J = 9 Hz, 4H), 8.18 (d, J = 9 Hz, 4H).
- (c) 2R,5R-Di-(4-nitrophenylsulfonyloxy)-1,6-diphenyl-3,4-O-isopropylidenehexane-3S,4S-diol (8.5 g) was dissolved in 300 mL of DMF and the solution was heated at 50 °C with about 9.2 g of NaN₃ and 6.3 g of 18-crown-6 for 4 h. The solvent was evaporated, the residue was taken up in ether and the mixture was extracted with aqueous NaHCO₃ solution. After washing with water, the organic phase was dried and concentrated. The residue was chromatographed on silica gel (toluene:n-heptane, 2:5 to 2:3). 2S,5S-Diazido-1,6-diphenyl-3,4-O-isopropylidenehexane-3R,4R-diol (2.37 g) was obtained. ¹H NMR (270 MHz, DMSO- d_6): 1.48 (s, 6H), 2.92-3.12 (m, 4H), 3.74 (dd, J = 10 Hz, 5 Hz, 2H), 4.15 (s, 2H), 7.21-7.39 (m, 10H).
- (d) 2S,5S-Diazido-1,6-diphenyl-1,6-O-isopropylidenehexane-3R,4R-diol (2.3 g) was dissolved in 50 mL of methanol and hydrogenated using about 0.2 g of palladium-on-charcoal (10%) under normal pressure for 2 h. The catalyst was filtered off, the solution was concentrated and the residue was chromatographed on silica gel (MC:ethanol, 99:1). The yield was 1.33 g. MS (FAB): 341 (M + H)⁺; ¹H NMR (270 MHz, DMSO- d_6): 1.29 (m, 4H), 1.37 (s, 6H), 2.71 (dd, J = 12 Hz, 5 Hz, 2H), 2.87 (m, 2H), 3.32 (m, 2H), 3.95 (s, 2H), 7.12–7.33 (m, 10H).
- N, N'-bis-(L-Valyl)-2S, 5S-diamino-1, 6-diphenylhexane-3R,4R-diol dihydrochloride (4). (a) 2S,5S-Diamino-1.6diphenyl-3,4-O-isopropylidenehexane-3R,4R-diol (136 mg) was dissolved in 2 mL of dry EA with 0.54 mL of NEM and 260 mg of N-tert-butoxycarbonyl-Lvaline. A 50% propanephosphonic anhydride (0.97 mL) solution in EA was added at -10 °C. The mixture was stirred at 0 °C for 1 h and then at rt overnight. The solution was diluted with EA and extracted with saturated NaHCO₃ solution, 10% KHSO₄ solution and water. The organic phase was dried over anhydrous MgSO₄ and concentrated and the residue was purified by chromatography on silica gel (methylene chloride: ethanol, 97:3). N,N-bis-(tert-Butoxycarbonyl-Lvalyl)-2S,5S-diamino-1,6-diphenyl-3,4-O-isopropylidene-

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hexane-3R,4R-diol (230 mg) was obtained. MS (FAB): 739 (M + H)⁺, 681, 639, 569, 539.

(b) N,N-bis-(tert-Butoxycarbonyl-L-valyl)-2S,5S-dia mino-1,6-diphenyl-3,4-O-isopropylidenehexane-3R,4R-diol (220 mg) was stirred into 10 mL of an approximately 3 N solution of HCl in dioxane: methanol, 1:1 at rt for 1 h. The volatile constituents of the solution were removed in vacuo and the residue was dried under a high vacuum. The substance was used without further purification. Yield: 184 mg; MS (FAB): 499 (M + H)⁺, 481, 463.

N, N'-bis-[(2S-(1,1-Dimethylethylsulfonylmethyl)-3-(1-Dimethyl)naphthyl)-propionyl)-L-valyl]-2S,5S-diamino-1,6-diphenylhexane-3R,4R-diol (6). N,N-bis-(L-Valyl)-2S,5Sdiamino-1,6-diphenylhexane-3R,4R-diol dihydrochloride 4 (57 mg), (2S-(1,1-dimethylethyl-sulfonyl-methyl)-3-(1-naphthyl)-propionic acid 5¹⁰ (95 mg), HOBt (41 mg) and TBTU (96 mg) were dissolved in 1 mL of dry DMF. N-ethyldiisopropylamine (0.11 mL) was added at rt and the mixture was stirred for 1 h. The solvent was evaporated, the residue was taken up in 30 mL of EA and the mixture was extracted with bisulfate solution, bicarbonate solution and water. After drying over Na₂SO₄, the extract was concentrated and the crude material was purified by chromatography on silica gel (MC:methanol, 97:3). The product was obtained in a yield of 31 mg. MS (FAB): 1153 (M + Na)⁺, 1131; ¹H NMR (270 MHz, DMSO- d_6): 0.69 (d, J = 7 Hz, 6H), 0.76 (d, J = 7 Hz, 6H), 1.10 (s, 18H), 1.86 (m, 2H),2.63-2.87 (m, 6H), 3.08 (m, 2H), about 3.25-3.44 (m, about 2H), 3.52-3.63 (m, 2H), 4.08 (m, 2H), 7.32 (d, J =8Hz, 2H), 7.38-7.48 (m, 4H), 7.47-7.62 (m, 4H), 7.81 (m, 2H), 7.92 (m, 2H), 8.12-8.25 (m, 4H).

1-Sulfoxy-2,5-dioxy-3R,4R-di-[1S-tert-butyloxycarbonyl-amino-2-phenylethyl]-cyclopentane (7). 2S,5S-Di-tert-butyloxycarbonylamino-1,6-diphenylhexane-3R,4R-diol). (300 mg, 0.6 mmol) was suspended in 10 mL MC and 1 mL absolute pyridine and treated at 0 °C with 86 mg (0.72 mol) SOCl₂. The mixture was diluted with methylenechloride, washed with brine and water, dried over Na₂SO₄, concentrated and chromatographed over silica (toluene:ethylacetate, 7:3). 320 mg (97%) of 7 were obtained. MS (FAB): 553.3 (M + Li⁺).

2S, 5S-Di-te rt-butyloxycarbonylamino-3R-hydroxy-4R-azido-1,6-diphenylhexane (8). Compound 7 (870 mg, 1.6 mmol) in 15 mL absolute DMF was treated with 311 mg (6.4 mmol) LiN₃ at 100 °C for 22 h. The solvent was removed by distillation, the product was dissolved in EA, washed with brine and water, dried over Na₂SO₄, concentrated and chromatographed over silica (toluene: EA 9:1). Compound 8 (286 mg) was obtained. MS (FAB): 531.8 (M + Li⁺). ¹H NMR (270 MHz, DMSO- d_6): 1.25 (s, 9H), 1.32 (s, 9H), 2.28 (d,2H), 2.54-2.68 (m, 2H), 3.57 (dd, 1H), 3.95-4.20 (m, 3H), 7.08 (d, 1H), 7.13-7.32 (m, 10H).

2S, 5S-Diamino-3R-hydroxy-4R-azido-1,6-diphenylhexane (9). Compound 8 (200 mg, 0.4 mmol) was dissolved in 10 mL 4.4 N HCl/dioxane and strirred at 0 °C for 1 h. The solvent was evaporated, the product was treated with diethylether, filtered and dried. Compound 9 (155 mg) was obtained and used without further purification.

N, N'-bis-[(2S-(1,1-Dimethylethylsulfonylmethyl)-3-(1-naphthyl)-propionyl)-L-valyl]-1,4-diaminobutane (10). Synthesis in analogy to 3: MS (FAB): 941 (M + Na)⁺, 919 (M + H)⁺; ¹H NMR (270 MHz, DMSO- d_6): 0.82 (d, J=6 Hz, 12H), 1.19 (s, 18H), 1.32 (m, 4H), 1.89 (m, 2H), 2.98 (m, 4H), 3.32 (m, 2H), 3.42 (m, 6H), 3.54 (dd, J=12.8 Hz, 8 Hz, 2H), 4.04 (t, J=8 Hz, 2H), 7.38 (m, 4H), 7.53 (m, 6H), 7.79 (m, 2H), 7.92 (m, 2H), 8.08 (d, J=8 Hz, 2H), 8.21 (m, 2H).

N, N'-bis-[(2S-(1,1-Dimethylethylsulfonylmethyl)-3-(1-naphthyl)-propionyl)-L-valyl]-1,4-diaminobutane-2R,3R-diol (11). Synthesis in analogy to 6 from 1,4-diaminobutane-2R,3R-diol dihydrochloride. MS (FAB): 973 (M + Na)⁺; 951 (M + H)⁺; 1 H NMR (270 MHz, DMSO- 4 6): 0.82 (d, 4 J = 6 Hz, 12H), 1.17 (s, 18H), 1.92 (m, 2H), 2.92-3.08 (m, 4H), 3.16-3.53 (m, 10H), 3.53 (dd, 4 J = 12.8 Hz, 8.8 Hz, 2H), 4.11 (dd, 4 J = 8.0 Hz, 7.2 Hz, 2H), 4.55 (d, 4 J = 4.8 Hz, 2H), 7.38-7.67 (m, 10H), 7.80 (m, 2H), 7.92 (m, 2H), 8.12 (d, 4 J = 8.4 Hz, 2H), 8.20 (d, 4 J = 8 Hz, 2H)

N, N'-bis-[(2S-(1,1-Dimethylethylsulfonylmethyl)-3-(1-naphthyl)-propionyl)-L-valyl]-2S, 5S-diamino-1, 6-diphenylhexane-3S, 4S-diol (12). Synthesis in analogy to 6; MS (FAB): 1131 (M + H)⁺, 716; ¹H NMR (270 MHz, DMSO- d_6): 0.77 (d, J = 7 Hz, 6H), 0.80 (d, J = 7 Hz, 6H), 1.12 (s, 18H), 1.87 (m, 2H), 2.75 (m, 2H), 2.83 (m, 2H), 2.92-3.03 (m, 2H), 3.10-3.22 (m, 2H), about 3.27-3.49 (m, 6H), 3.54-3.67 (m, 2H), 4.02-4.15 (m, 4H), 4.66 (d, J = 6 Hz, 2H), 7.01-7.09 (m, 2H), 7.10-7.25 (m, 8H), 7.28-7.43 (m, 4H), 7.48-7.68 (m, 6H), 7.79 (d, J = 8 Hz, 2H), 7.88-7.95 (m, 2H), 8.15-8.25 (m, 4H).

N, N'-bis- $[(2S-(1,1-Dimethylethylsulfonylmethyl)-3-(1-naphthyl)-propionyl)-L-valyl]-2S,5S-diamino-1,6-diphenylhexane-3R,4S-diol (13). Synthesis in analogy to 6. MS (FAB): 1153 (M + Na)<math>^+$, 1131.

N, N'-bis-[(2S-(1,1-Dimethylethylsulfonylmethyl)-3-(1-naphthyl)-propionyl)-L-valyl]-2S,5S-diamino-1,6-diphenyl-3R-hydroxy-4S-azidohexane (14). Synthesis in analogy to 6 from 9. MS (FAB): 1162.6 (M + Li)⁺, 1156.6.

N, N'-bis-[(2S-(1,1-Dimethylethylsulfonylmethyl)-3-(1-naphthyl)-propionyl)-L-valyl]-2S,5S-diamino-1,6-diphenyl-3R-hydroxy-4S-aminohexane (15). Compound 14 (89 mg, 0.08 mmol) was dissolved in 15 mL methanol and hydrogenated over 100 mg Pd/C (10%), 1 atm for 12 h. The catalyst was filtrated, the solvent was evaporated and the residue chromatographed over silica (MC:methanol, 9:1) and 36 mg (41%) of 15 was obtained. MS (FAB): 1136.7 (M + Li)⁺, 1130.7.

N, N'-bis-[(2S-(1,1-Dimethylethylsulfonylmethyl)-3-(1-naphthyl)-propionyl)-L-valyl]-2S, 5S-diamino-1,6-diphenyl-3S-hydroxyhexane (16). Synthesis in analogy to 6 starting from 3S-hydroxy-2S, 5S-diamino-1,6-diphenylhexane. MS (FAB): 1137 (M + Na)+, 1115.

N, N'-bis-([2S-(1,1-Dimethylethylsulfonylmethyl)-3-(1-naphthyl)-propionyl]-L-valyl)-3S,6S-diamino-1,8-diphenyloctane-4R,5R-diol (17). Synthesis in analogy to 6 from 3S,6S-diamino-1,8-diphenyloctane-4R,5R-diol dihydrochloride (the latter compound was prepared in analogy to 3 from 2 and benzyllithium). MS (FAB(LiI)): 1165 (M + Li)⁺; ¹H NMR (270 MHz, DMSO- d_6): 0.92 (d, J = 7 Hz, 12H), 1.13 (s, 18H), 1.6-1.85 (m, 4H), 2.04 (m, 2H), 2.40-2.64 (m, 4H), 2.82 (dm, J = 14 Hz, 2H), 3.18 (m, 2H), 3.32-3.52 (m, 6H), 3.58 (m, 2H), 4.08 (m, 2H), 4.22 (t, J = 8 Hz, 2H), 7.1-7.56 (m, 20H), 7.72 (dd, J = 4 Hz, 2H), 7.88 (m, 2H), 8.14 (m, 2H), 8.32 (d, J = 8 Hz, 2H).

N, N'-bis-[(2S-(1,1-Dimethylethylsulfonylmethyl)-3-(1-naphthyl)-propionyl)-L-valyl]-2S,5S-diamino-1,6-di-(4-hydroxyphenyl)-hexane-3R,4R-diol (18). Synthesis in analogy to 6 from 2S,5S-diamino-1,6-bis-(4-hydroxyphenyl)-hexane-3R,4R-diol dihydrochloride. MS (FAB): 1185 (M + Na)⁺, 1163.

N, N'-bis-([2S-1, 1-Dimethylethylsulfonylmethyl)-3-(1naphthyl)-propionyl]-L-valyl)-2S,5S-diamino-1,6-bis-(3,4-methylenedioxyphenyl)-hexane-3R,4R-diol Synthesis in analogy to 6 from 2S,5S-diamino-1,6-bis-(3,4-methylenedioxyphenyl)-hexane-3R,4R-diol dihydrochloride (the latter compound was prepared in analogy to 3 from 2 and 3,4-methylenedioxyphenyllithium). MS (FAB): $1241 (M + Na)^+$, $1219 (M + H)^+$; ¹H NMR (270) MHz, DMSO- d_6): 0.73 (d, J = 7 Hz, 6H), 0.78 (d, J = 7Hz, 6H), 1.10 (s, 18H), 1.89 (m, 2H), 2.55-2.72 (m, 4H), 2.79 (dm, J = 14 Hz, 2H), 3.08 (dd, J = 14 Hz, 10 Hz, 2H), about 3.22-3.43 (m, about 6H), 3.58 (dd, J =14 Hz, 10 Hz, 2H), 4.07 (m, 2H), 4.45 (m, 2H), 4.49 (m, 2H), 5.75 (s, 2H), 5.78 (s, 2H), 6.68 (s, 2H), 6.80 (s, 2H), 7.25 (d, J = 9 Hz, 2H), 7.39-7.45 (m, 4H), 7.54 (m, 6H), 7.80 (m, 2H), 7.92 (m, 2H), 8.15-8.25 (m, 4H).

N, N'-bis-([2S-(1,1-Dimethylethylsulfonylmethyl)-3-(1-naphthyl)-propionyl]-L-valyl)-2S,5S-diamino-1,6-bis-(2,4-dimethoxyphenyl)-hexane-3R,4R-diol (20). Synthesis in analogy to 3 from 2S,5S-diamino-1,6-bis-(2,4-dimethoxyphenyl)-hexane-3R,4R-diol dihydrochloride (the latter compound was prepared in analogy to 3 from 2 and 2,4-dimethoxyphenyllithium). MS (FAB): $1251 (M + H)^+$.

N, N'-bis-[(2S-(1,1-Dimethylethylsulfonylmethyl)-3-(1-naphthyl)-propionyl)-L-valyl]-3S,6S-diamino-1, 8-di-(4-pyridyl)-octane-4R,5R-diol (21). Synthesis in analogy to 6 from 3S,6S-diamino-1,8-di-(4-pyridyl)-octane-4R,5R-diol-tetrahydrochloride (the latter compound was obtained in analogy to 3 starting from 2 and 4-picolyllithium. MS (FAB): 1161 (M + H)+; 1 H NMR (270 MHz, DMSO- d_6): 0.83 (m, 12H), 1.14 (s, 18H), 1.66 (m, 2H), 1.82 (m, 2H), 2.00 (m, 2H), 2.50-2.78 (m,

4H), 2.86 (m, 2H), 3.06–3.63 (m, 10H), 4.02 (m, 2H), 4.14 (m, 2H), 4.69 (m, 2H), 7.30-7.60 (m, 14H), 7.74 (d, J = 8 Hz, 2H), 7.87 (m, 2H), 8.16 (m, 2H), 8.32 (d, J = 8 Hz, 2H), 8.58 (m, 4H)

N, N'-bis-([2S-(1,1-Dimethylethylsulfonylmethyl)-3-(1naphthyl)-propionyl]-L-valyl)-6S,9S-diaminotetradecane-7R,8R-diol (22). Synthesis analogous to 6 from 6S.9S-diaminotetradecane-7R,8R-diol dihydrochloride (the latter compound was prepared in analogy from 2 and n-butyllithium). MS (FAB(LiI)): 1097 (M + Li)⁺; ¹H NMR (270 MHz, DMSO- d_6): 0.76 (m, 6H), 0.88 (d, J = 7 Hz, 12H), 1.12 (s, 18H), about 1.10-1.54(m, 16H), 2.02 (m, 2H), 2.82 (dd, J = 12 Hz, 2 Hz, 2H),3.16 (dd, J = 12 Hz, 16 Hz, 2H), 3.24 (m, 2H), 3.363.52 (m, 4H), 3.58 (dd, J = 8 Hz, 13 Hz, 2H), 3.98 (m, 4H)2H), 4.16 (t, J = 6 Hz, 2H), 4.44 (s, 2H), 7.18 (d, J = 10Hz, 2H), 7.42-7.48 (m, 4H), 7.49-7.62 (m, 4H), 7.81 (m, 2H), 7.92 (m, 2H), 8.20 (d, J = 8 Hz, 2H), 8.30 (d, J =8.4 Hz, 2H).

N, N'-bis-([2S-(1,1-Dimethylethylsulfonylmethyl)-3-(1-naphthyl)-propionyl]-L-isoleucyl)-2S,5S-diamino-1,6-diphenylhexane-3R,4R-diol (23). Synthesis in analogy to 6. MS (FAB): 1181 (M + Na)⁺; ¹H NMR (270 MHz, DMSO- d_6): 0.63 (d, J=7 Hz, 6H), 0.73 (t, J=7 Hz, 6H), 0.99 (m, 2H), 1.11 (s, 18H), 1.32 (m, 2H), 1.64 (m, 2H), 2.63–2.88 (m, 6H), 3.07 (dd, J=15 Hz, 11 Hz, 2H), about 3.28–3.43 (m, about 6H), 3.58 (dd, J=14 Hz, 9 Hz, 2H), 4.09 (t, J=8 Hz, 2H), 4.48–4.62 (m, 4H), 7.03 (m, 2H), 7.12–7.31 (m, 10H), 7.43 (m, 4H), 7.54 (m, 4H), 7.81 (m, 2H), 7.92 (m, 2H), 8.15–8.25 (m, 4H).

N, N'-bis-[(2S-(1, 1-Dime thylethylsulfonylmethyl)-3-phenyl-propionyl)-L-tert-butylglycyl]-2S,5S-diamino-I,6-diphenylhexane-3R,4R-diol (24). Synthesis in analogy to 6. MS (FAB): 1181 (M + Na)⁺, 1159 (M + H)⁺; ¹H NMR (270 MHz, DMSO- d_6): 0.83 (s, 18H), 1.12 (s, 18H), 2.39 (dd, J=11 Hz, 14 Hz, 2H), 2.56–2.72 (m, 4H), 2.73–2.90 (m, 4H), about 3.25–3.40 (m, about 4H), 3.53 (dd, J=10 Hz, 14 Hz, 2H), 4.20 (d, J=9 Hz, 2H), 4.54 (m, 2H), 4.62 (m, 2H), 6.98 (m, 2H): 7.07–7.36 (m, 18H), 7.37 (d, J=9 Hz, 2H), 7.98 (d, J=9 Hz, 2H).

N, N'-bis-[(2S-(1,1-Dimethylethylsulfonylmethyl)-3-(1-naphthyl)-propionyl)-L-cyclohexylglycyl]-2S, 5S-diamino-1,6-diphenylhexane-3R,4R-diol (25). Synthesis in analogy to 6. MS (FAB): 1233 (M + Na) $^+$, 1211 (M + H) $^+$.

N, N'-bis-[(2S-(1,1-Dimethylethylsulfonylmethyl)-3-(1-naphthyl)-propionyl)-L-cyclopentylglycyl]-2S, 5S-diamino-1,6-diphenylhexane-3R,4R-diol (26). Synthesis in analogy to 6. MS (FAB): 1205 (M + Na)⁺, 1183 (M + H)⁺.

N, N'-bis-[(2S-(1,1-Dimethylethylsulfonylmethyl)-3-(1-naphthyl)-propionyl)-L-propylglycyl]-2S, 5S-diamino-1, 6-diphenylhexane-3R, 4R-diol (27). Synthesis in analogy to 6. MS (FAB): 1153 (M + Na)^+ , 1131 (M + H)^+ .

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N, N'-bis- $[(2S-(1,1-Dimethylethylsulfonylmethyl)-3-(1-naphthyl)-propionyl)-homoglycyl]-2S, 5S-diamino-1, 6-diphenylhexane-3R, 4R-diol (28). Synthesis in analogy to 6. MS (FAB): <math>1097 (M + Na)^+$, $1075 (M + H)^+$.

N, N'-bis-([2S-(1,1-Dimethylethylsulfonylmethyl)-3-(1-naphthyl)-propionyl]-L-asparaginyl)-2S,5S-diamino-1, 6-diphenylhexane-3R,4R-diol (29). Synthesis in analogy to 6. MS (FAB): 1183 (M + Na)⁺; ¹H NMR (270 MHz, DMSO- d_6): 1.17 (s, 18H), 2.22 (m, 2H): 2.37-2.76 (m, 10H), 2.90 (m, 2H), 3.25 (m, 4H), 3.58 (m, 2H), 4.25 (m, 2H), 4.40 (m, 2H), 4.62 (m, 2H), 6.93-7.60 (m, 24H), 7.77 (m, 2H), 7.90 (m, 2H), 8.22 (d, J = 8 Hz, 2H), 8.33 (d, J = 8 Hz, 2H).

N, N'-bis-([2S-(1,1-Dimethylethylsulfonylmethyl)-3-(1-naphthyl)-propionyl]-L-threonyl)-2S, 5S-diamino-1, 6-diphenylhexane-3R, 4R-diol (30). Synthesis in analogy to 6. MS (FAB): 1157 (M + Na)^+ , 1135 (M + H)^+ .

N, N'-bis-([2S-(1,1-Dimethylethylsulfonylmethyl)-3-(1-naphthyl)-propionyl]-L-glutaryl)-2S,5S-diamino-1,6-diphenylhexane-3R,4R-diol (31). Synthesis in analogy to 6. MS (FAB): $1213 (M + Na)^+$, $1191 (M + H)^+$.

N, N'-bis-([2S-(1,1-Dimethylethylsulfonylmethyl)-3-(1-naphthyl)-propionyl]-L-histidyl-2S, 5S-diamino-1, 6-diphenylhexane-3R, 4R-diol (32). Synthesis in analogy to 6. MS (FAB): $1229 (M + Na)^+$, $1207 (M + H)^+$.

N, N'-bis-([2S-(1,1-Dimethylethylsulfonylmethyl)-3-(1-naphthyl)-propionyl]-L-prolyl)-2S,5S-diamino-1,6-diphenylhexane-3R,4R-diol (33). Synthesis in analogy to 6. MS (FAB): 1149 (M + Na)^+ , 1127 (M + H)^+ .

N, N'-bis-(L-Phenylalanyl-L-valyl)-2S,5S-diamino-1,6-diphenylhexane-3R,4R-diol dihydrochloride (34). Compound 4 (100 mg) was dissolved in DMF (1.5 mL) together with N-tert-butoxycarbonyl-L-phenylalanine (111 mg), NEM (0.57 mL) and HOBt (60 mg). After addition of EDAC (85 mg) at 0 °C, stirring was continued at 0 °C for 1 h and then at rt overnight. The solvent was evaporated, the residue was taken up in EA and the mixture was extracted with saturated KHCO3 solution, 10% KHSO₄ solution and water. The organic was dried with anhydrous Na₂SO₄ concentrated. The residue was recrystallized from ethanol-water. The yield of N,N-bis-(tert-butoxycarbonyl-L-phenylalanyl-L-valyl)-2S, 5S-dia mino-1, 6diphenylhexane-3R,4R-diol was 92 mg. MS (FAB): 993 $(M + H)^+$, 975, 893, 793; ¹H NMR (270 MHz, DMSO d_6): 0.72 (d, J = 6 Hz, 6H), 0.75 (d, J = 6 Hz, 6H), 1.29 (s, 18H), 1.86 (m, 2H), 2.60-2.96 (m, 8H), 3.30 (m,2H), 4.17 (m, 2H), 4.45 (m, 2H), 4.68 (m, 2H), 7.03 (d, J = 9 Hz, 2H), 7.05–7.30 (m, 22H), 7.53 (d, J = 9 Hz,

(b) N,N-bis-(tert-Butyloxycarbonyl-L-phenylalanyl-L-valyl)-2S,5S-diamino-1,6-diphenylhexane-3R,4R-diol (80 mg) was treated with a mixture of 2 mL of 5 N HCl in dioxane and 1 mL of HCl in methanol at rt for 30 min. The volatile constituents were removed in vacuo, the residue was washed with ether and the substance

was dried under a high vacuum. The yield was 50 mg; MS (FAB): 793 (M + H)⁺, 775.

N, N'-bis-[(3-Amino-2-benzyl)-propionyl-L-valyl]-2S,5S-diamino-1,6-diphenylhexane-3R,4R-diol dihydrochloride (35). Synthesis in analogy to 34 from 4 and 2-benzyl-3-tert-butoxycarbonylaminopropionic acid. MS (FAB): 843 (M + Na)⁺, 821 (M + H)⁺.

N, N'-bis-[(2R-Benzyl-3-carboxyl)-propionyl-L-valyl]-2S.5S-diamino-1.6-diphenylhexane-3R.4R-diol (36). (a) Compound 4 (45 mg) was dissolved in DMF (2 mL) with 2R-benzyl-3-tert-butoxycarbonylpropionic acid (75 mg) (prepared after lit. 17), and HOBt (37 mg), TBTU (87 mg) and ethyldiisopropylamine (112 μL) were added. The mixture was stirred at rt for 15 min, the DMF was removed in vacuo, the residue was taken up in EA and the mixture was extracted with KHSO4 solution, NaHCO3 solution and water. The organic phase was dried over MgSO₄ and concentrated. The residue was triturated with diethyl ether and filtered off. N,N-bis-[(2R-Benzyl-3-tert-butoxycarbonyl)-prop-ionyl-L-valyl]-2S,5S-diamino-1,6-diphenylhexane-3R,4R-diol (44 mg) was obtained. MS (FAB): $1013 (M + Na)^+$ 991 (M + H)⁺; ¹H NMR (270 MHz, DMSO- d_6): 0.69 (d, J = 6 Hz, 6H), 0.74 (d, J = 6 Hz, 6H), 1.31 (s, 18H), 1.83 (m, 2H), 1.95 (m, 2H), 2.32-2.47 (m, 4H), 2.60-2.87 (m, 6H), 2.98 (m, 2H), 3.29 (m, 2H), 4.09 (dd, J =8 Hz, 7 Hz, 2H), 4.46 (m, 2H), 4.64 (m, 2H), 7.02-7.31 (m, 10H), 7.38 (d, J = 9 Hz, 2H), 7.80 (d, J = 8 Hz,2H).

(b) N,N-bis-[(2R-benzyl-3-carboxyl)-propionyl-L-valyl]-2S,5S-diamino-1,6-diphenylhexane-3R,4R-diol **36** (15 mg) was obtained from N,N-bis-[(2R-benzyl-3-tert-butoxyarbonyl)-propionyl-L-valyl]-2S,5S-diamino-1,6-diphenylhexane-3R,4R-diol (30 mg) by treatment with trifluoroacetic acid. MS (FAB): 901 (M + Na)⁺, 879 (M + H)⁺.

N, N'-bis-[(2S-Hydroxy-3-phenylpropionyl)-L-valyl]-2S.5S-diamino-1.6-diphenylhexane-3S.4S-diol (37). HOBt (27 mg), of TBTU (64 mg) and then, slowly, diisopropylethylamine (0.088 mL) were added to N,N'bis-[-L-valyl]-2S,5S-diamino-1,6-diphenylhexane-3S,4Sdiol dihydrochloride (0.065 mmol) and S-phenyllactic acid (33 mg) in DMF (4 mL). After 15 min at rt, the DMF was removed in vacuo, the residue was taken up in EA and the mixture was extracted with KHSO₄ solution, NaHCO₃ solution and water. The organic phase was dried with MgSO₄ and concentrated, and the residue was triturated with ether and filtered. The yield was 43 mg. MS (FAB): 795 (M + H)⁺; ¹H NMR (270 MHz, DMSO- d_6): 0.63 (d, J = 7 Hz, 6H), 0.67 (d, J = 7Hz, 6H), 1.82 (m, 2H), 2.64-2.79 (m, 4H), 2.91-3.04 (m, 4H), 3.38 (m, 2H), 3.97-4.17 (m, 6H), 4.72 (d, J = 6 Hz, 2H), 5.77 (d, J = 6 Hz, 2H), 7.08-7.29 (m, 20H), 7.38 (d, J = 9 Hz, 2H), 7.85 (d, J = 8 Hz, 2H).

N, N'-bis-([1-Naphthyloxy]-acetyl-L-valyl)-2S, 5S-diamino-1, 6-diphenylhexane-3R, 4R-diol (41). Synthesis analogous to 6 from 4 and 1-naphthyloxyacetic acid. MS (FAB): $866 (M + H)^+$.

N, N'-bis-[D-Gluconyl-L-phenylalanyl-L-valyl]-2S,5S-diamino-1,6-diphenylhexane-3R,4R-diol (42). (a) N,N'-bis-[2,3,4,5,6-Penta-O-acetyl-D-gluconyl-L-phenylalanyl-L-valyl]-2S,5S-diamino-1,6-diphenylhexane-3R,4R-diol. Synthesis by coupling of 2,3,4,5,6-penta-O-acetyl-D-gluconic acid¹⁸ and 34 in analogy to 6. MS (FAB): 1569 (M + H)⁺.

(b) Compound 42 was obtained from N,N-bis-[2,3,4,5,6-penta-O-acetyl-D-gluconyl-L-phenylalanyl-L-valyl]-2S,5S-diamino-1,6-diphenylhexane-3R,4R-diol with a saturated solution of ammonia in methanol. MS (FAB): 1171 (M + Na) $^+$.

N, N'-bis-[(2S-(1,1-Dimethylethylsulfonylmethyl)-3-phenylpropionyl)-L-valyl]-2S,5S-diamino-1,6-diphenylhexane-3R,4R-diol (43). Synthesis in analogy to 6. MS (FAB): 1053 (M + Na)⁺, 1031 (M + H)⁺; ¹H NMR (270 MHz, DMSO- d_6): 0.72 (d, J=7 Hz, 6H), 0.78 (d, J=7 Hz, 6H), 1.14 (s, 18H), 1.85 (m, 2H), 2.62-2.94 (m, 8H), about 3.20-3.35 (m, about 4H), 3.53 (dd, J=10 Hz, 14 Hz, 2H), 4.02-4.13 (m, 2H), 4.50 (m, 2H), 4.64 (m, 2H), 7.01-7.10 (m, 2H), 7.12-7.39 (m, 22H), 8.05 (J=8 Hz, 2H).

N, N'-bis-[(2S-(1,1-Dimethylethylsulfonylmethyl)-3-(4-methoxyphenyl)propionyl)-L-valyl]-2S,5S-diamino-1,6-diphenylhexane-<math>3R,4R-diol (44). Synthesis in analogy to 6. MS (FAB): 1113 (M + Na)⁺, 1091 (M + H)⁺.

N, N'-bis-([2S-(1,1-Dimethylethylsulfonylmethyl)-3-(4-pyridyl)-propionyl]-L-valyl)-2S, 5S-diamino-1, 6-diphenylhexane-3R, 4R-diol dihydrochloride (45). Synthesis in analogy to 6; 2S-(1,1-dimethylethylsulfonylmethyl)-3-(4-pyridyl)-propionic acid is synthesized in analogy to Ref. 10. MS (FAB): 1055 (M + Na)⁺, 1033 (M + H)⁺; H NMR (270 MHz, DMSO- d_6): 0.68 (d_1 , d_2 , d_3 , d_4 , d_4 , d_5 , d_5 , d_5 , d_6 , d_7 , d_7 , d_7 , d_8 , d

N, N'-bis-([2-(1,1-Dimethylethylsulfonylmethyl)-3-(N-oxido-4-pyridyl)-propionyl]-L-valyl)-2S, SS-diamino-1, 6-diphenylhexane-3R, 4R-diol (46). Synthesis in analogy to 6. 2S-(1,1-Dimethylethylsulfonylmethyl)-3-(N-oxido-4-pyridyl)-propionic acid is formed from 2S-(1,1-dimethylethylsulfonylmethyl)-3-(4-pyridyl)-propionic acid by oxidation with an additional equivalent of oxone. MS (FAB): 1065 (M + H)⁺.

N, N'-bis-[(3-(1,1-Dimethylethylsulfonyl)-propionyl)-L-valyl]-2S, 5S-diamino-1,6-diphenylhexane-3R, 4R-diol (47). Synthesis in analogy to 6. MS (FAB): 873 (M + Na)⁺, 851 (M + H)⁺; ¹H NMR (270 MHz, DMSO- d_6): 0.69 (d, J = 6 Hz, 6H), 0.73 (d, J = 6 Hz, 6H), 1.33 (s, 18H), 1.84 (m, 2H), 2.54-2.59 (m, 6H), 2.67 (m, 2H), about 3.15-3.30 (m, 6H), 4.05 (dd, J = 7 Hz, 9 Hz, 2H), 4.47 (m, 2H), 4.63 (m, 2H), 7.06-7.21 (m, 10H), 7.30 (d, J = 9 Hz, 2H), 7.94 (d, J = 8 Hz, 2H).

N, N'-bis-[(2-(2-Hydroxyethylsulfonylmethyl)-3-phenyl-propionyl)-L-valyl]-2S, 5S-diamino-1, 6-diphenylhexane-3S, 4S-diol (48). Synthesis in analogy to 6. MS (FAB): 1007 (M + H)⁺.

N,N'-bis-[(2-Decylsulfonylmethyl)-3-phenylpropionyl)-L-valyl]-2S,5S-diamino-1,6-diphenylhexane-3S,4S-diol (49). Synthesis in analogy to 6. MS (FAB): 1221 (M + Na)⁺, 1199 (M + H)⁺.

N, N'-bis-(2-[4-Pyridyl]ethylsulfonyl-L-valyl)-2S,5S-diamino-1,6-diphenylhexane-3R,4R-diol (50). Synthesis in analogy to 6. MS (FAB): 836 (M + H)⁺.

N, N'-bis-[(2-Methylsulfonylmethyl)-3-phenylpropionyl)-L-valyl]-2S,5S-diamino-1,6-diphenylhexane-3S,4S-diol (51). Synthesis in analogy to 6 and 34. MS (FAB): 1069 (M+Na)⁺, 1047 (M+H)⁺.

Biology. HIV Protease inhibition assay

Recombinant HIV-1 protease was expressed and purified from *Escherichia coli* strain K12 as described.¹²

Inhibition of HIV protease was determined using an HPLC-test with the synthetic substrate SFNFOprQI, where Opr stands for 5-oxaproline and substitutes the proline in the natural sequence SFNFPQI (N-terminus of the protease). The Opr-containing peptide is a much better substrate than the natural one and allows the reduction of the amount of enzyme necessary for cleavage.7 The resulting tetrapeptide fragment H-Ser-Phe-Asn-Phe-OH is easily separated and quantitated by reverse phase HPLC. The HIV protease activity was measured at 37 °C in 0.4 M sodium phosphate, pH 5.6, containing 20 mM morpholinoethanesulfonic acid, 10% (w/v) glycerol, 0.01% (v/v) Triton X 100, 0.9 mM substrate and 5 mM EDTA in a total volume of 25 µL. The protease concentration was 0.5 nM. Inhibitors were added to the assay in dimethylsulfoxide (1 µL, final DMSO concentration = 4%) The reaction was stopped after 2 h by the addition of 175 µL HPLC solvent mixture.

Assay dilution (20 μ L) was analyzed using a Radial-Pak Liquid Chromatography Cartridge (Waters 8C18/5 μ) by isocratic elution with a solution containing 78% (w) 0.1 M H₃PO₄, pH 2.5 with triethylamine, and 22% (w) acetonitrile at a flow rate of 1 mL min⁻¹. Peaks were detected by UV absorbance at 215 nm, retention times were 5 min for the tetrapeptide and 9 min for the substrate.

Inhibition of HIV-1 viral spread in cell culture

Stock solutions of inhibitors were prepared by dissolving 2 mg of inhibitor in 120 µL DMSO.

Virus stocks. Infectious HIV was obtained from supernatant fluids of infected peripheral blood mononuclear cells (PBL) and stored frozen in aliquots at -80 °C. HIV-1 (D34) is a particularly fast growing

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HIV-1 strain of the subtype B¹⁴ isolated from a German AIDS patient in 1985. ^{15b}

Cells. Peripheral blood mononuclear cells (PBL) from healthy HIV seronegative volunteers were isolated by Ficoll-Hypaque sedimentation of citrate containing blood. Cells were stimulated with phythaemagglutinine (PHA) and grown in RPMI-1640 supplemented with 20% fetal calf serum (FCS), antibiotics, HEPES and additional glutamine at 37 °C. Two days later 40 units of IL-2 (interleukin-2; Roussel Uclaf, France) were added per mL cell suspension. Stimulated cells were stored in liquid nitrogen.

Antiviral drug assay. PBL or H9 cells15b were cultivated for three to four days in RPMI-1640 supplemented with 20% FCS and (in the case of PBL cells) 40 IU mL⁻¹ recombinant IL-2, then they were infected for a 30 min period with HIV-1 (2-4 \times 10⁵ TCIU/10⁶ cells, TCIU: T cell infective units). The virus containing supernatant was removed, 0.5 mL of the resuspended infected cells were transferred immediately to the wells of a 24-well plate each containing 0.5 mL well⁻¹ of test compounds dissolved in DMSO. Dilutions of test compounds were prepared by serial dilutions of 1:5 or 1:2. The final concentration of the cells was $1-2.5 \times 10^5$ ml⁻¹ well⁻¹. The samples were incubated at 37 °C for 72-96 h under 5% CO₂. Virus replication was evaluated by light microscopy for syncytia formation and/or measurement of HIV antigen in the supernatant of the cell cultures by an antigen capture assay (Vironostika HIV, Organon Teknika).

Antigen capture assay. As antigen capture assay the ELISA assay "Vironostika HIV" (Organon Teknika, Eppelheim, Germany) was used. The main, but not exclusive, recognized antigen was the HIV p24 antigen. All assay procedures were performed according to the instructions of the manufacturer.

Cut-off values were defined by adding three standard deviations to the mean extinction values of negative controls. For the calculation of the IC_{50} values the supernatants of the corresponding cell cultures were tested in a 1:10 dilution in the antigen capture assay for the HIV-1 strain.

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